

### **COMMENTARY**

## Therapy by Taking Away: The Case of Iron

Barbara S. Polla\*

LABORATOIRE DE PHYSIOLOGIE RESPIRATOIRE, UFR COCHIN PORT-ROYAL, 75014 PARIS, FRANCE

ABSTRACT. The recent finding of the beneficial effects of iron deprivation in the outcome of muscle necrosis in an animal model of genetic myopathy served as the basis of this commentary. Here, "taking away" iron by controlled dietary deprivation is proposed as a reasonable, feasible, cheap, and efficient clinical approach to many diverse diseases, all of which have a free radical component. Indeed, iron potentiates the generation of the highly reactive and toxic hydroxyl radical, and, thus, of oxidative damage. Iron deprivation may represent the first really efficient antioxidant, preventing oxidative stress in all subcellular compartments, tissues, and organs. Iron/iron deprivation also modulates programmed cell death (apoptosis), which should be the subject of further studies to better define the mechanisms mediating these complex effects. Finally, related to its antioxidant effects, iron deprivation may find applications in the anti-aging field, whether programmed or premature aging, and whether in cosmetics or in gerontology. BIOCHEM PHARMACOL 57;12:1345–1349, 1999. © 1999 Elsevier Science Inc.

**KEY WORDS.** reactive oxygen species; iron; iron chelators; Fenton reaction; ischemia-reperfusion; infectious diseases; aging

Classical pharmacotherapy usually ends by giving the patient some magic in the form of a lively, colored, modernly shaped, and shiny pill: something was missing, something good is being administered. Here we propose that many diseases as diverse as myocardial infarction or stroke, inflammatory conditions such as bleomycin-induced pulmonary fibrosis, bacterial infections and malaria, or DMD†, all of which have a free radical component, would actually benefit most from taking away—in this case, iron. Iron deprivation appears to be more efficient and more feasible than any classical antioxidant therapy. Indeed, the latter has been limited by the heterogeneity of the clinical courses of the many diseases in which oxidative stress plays a role, and by homeostatic mechanisms that complicate the specificity of tissue and subcellular targeting of antioxidants. Thus, alternative approaches such as dietary deprivation or iron chelation are being considered as attractive therapeutic avenues, more likely to provide both efficient and safe antioxidant protection than any scavengers, vitamins, or trace elements.

### ROS, IRON, AND FERRITIN

ROS include the free radicals superoxide  $(O_2^{\bullet-})$  and hydroxyl ( ${}^{\bullet}OH$ ), the non-radical intermediates singlet

oxygen ( $^{1}O_{2}$ ) and hydrogen peroxide ( $H_{2}O_{2}$ ), as well as nitric oxide ( $^{\bullet}NO$ ) and peroxynitrites ( $ONOO^{-}$ ). At low concentrations, ROS can act as second messengers, gene regulators, and mediators for cell activation. But under conditions of excessive production or reduced availability or efficiency of antioxidant mechanisms, ROS may accumulate to unbalanced levels and may then damage almost all cellular components, whether proteins (unfolding, aggregation, desolubilization, and loss of function), lipids (peroxidation of cell and subcellular membranes), DNA, or mitochondria.

Iron, although indispensable to the function of many proteins, including enzymes involved in fundamental processes such as mitochondrial respiration, plays a major role in oxidative stress via Fenton chemistry, where iron(II) is stoichiometrically oxidized by H<sub>2</sub>O<sub>2</sub> to iron(III), producing OH [1]. Thus, among the ROS, the highly reactive and extremely toxic \*OH is produced whenever (and wherever) iron is available for the Haber-Weiss reaction (Fe<sup>3+</sup> +  $O_2^{\bullet-} \gg Fe^{2+} + O_2$ ; Fe<sup>2+</sup> +  $H_2O_2 \gg Fe^{3+} + O_1^{\bullet} + O_2^{\bullet} = 0$ OH-), which is particularly the case in the vicinity of DNA or in the mitochondria; indeed, normal cellular respiration is associated with the production of  $O_2^{\bullet-}$  and  $H_2O_2$ , thus providing the substrates, in the presence of iron, for \*OH generation. Furthermore, iron contributes to redox cycling of cytotoxic compounds such as Adriamycin<sup>®</sup> or bleomycin, which have a high affinity for DNA. The central role of iron in oxygen toxicity and the ability of iron deprivation to prevent \*OH formation by inhibiting the iron-driven Haber-Weiss reaction provides a strong rationale for the use of iron chelators or iron-deficient diets in the prevention of oxidative stress.

<sup>\*</sup> Correspondence: Dr. Barbara S. Polla, Laboratoire de Physiologie Respiratoire, UFR Cochin Port-Royal, 24 rue du Faubourg Saint-Jacques, 75014 Paris, France. Tel. 33-1-44-41-23-36; FAX 33-1-44-41-23-38. E-mail: Barbara.Polla@cochin.univ-paris5.fr

<sup>†</sup>Abbreviations: DMD, Duchenne muscular dystrophy; ROS, reactive oxygen species; IRE, iron-responsive element; IRP, iron regulatory protein; and TNF, tumor necrosis factor.

1346 B. S. Polla

Endogenously, the availability of iron is tightly regulated within mammalian cells, in particular by the expression of ferritin [2, 3]. Ferritin plays a central role in the cellular protection mechanisms against iron-mediated oxidative injury, and it has been proposed that ferritin should be considered as part of the stress protein families [4]. As other stress proteins, ferritin is ubiquitous and highly conserved. Ferritin specifically binds—"chaperones"—iron, thus protecting cells from the deleterious consequences of ironcatalyzed free radical reactions. Ferritin expression is regulated post-transcriptionally by an iron-dependent mechanism consisting of the cis-acting IRE and trans-acting IRE binding proteins, IRP1 and IRP2 [2, 3]. In iron-replete cells, IRP1 fulfills the role of a cytosolic aconitase bound to a 4Fe-4S cluster, while in iron-depleted cells, IRP1 is associated with ferritin mRNA, repressing the translation of ferritin. Transition between the two forms is regulated by the redox status of the cluster. Interestingly, it has been suggested recently that at least part of the beneficial effects of low-dose aspirin in the prevention of atherosclerosis, one of the key manifestations of oxidative stress in humans, could be related to an increase in ferritin synthesis in endothelial cells [5].

#### IRON AND APOPTOSIS

Apoptosis, from the ancient Greek "falling off" (the tree leaves), is a sophisticated system for programmed cell death and is involved in a number of fundamental biological processes such as metamorphosis, embryonic morphogenesis, development, and removal of superfluous, damaged, or transformed cells. Apoptosis occurs as a counterpart of proliferation in growth regulation and has been associated with pathological processes such as cancer, infections, and chronic inflammation [6–8]. Apoptosis can be induced by a large variety of agents, but, regardless of the inducing agent, the apoptotic process is usually divided into three phases: induction, control, and execution/degradation, all of which have distinct biochemical and morphological features [9–11].

The role of iron in apoptosis remains controversial. In particular, the relationships between ROS, iron, and apoptosis are complex. On the one hand, iron contributes to oxidative stress, a well-characterized inducer of apoptosis [12], and, indeed, iron has, for example, been shown to accelerate endothelial cell apoptosis via a ROS-dependent mechanism [13]. On the other hand, iron is essential for cell growth and viability, and leads to an increased expression of antiapoptotic proteins of the Bcl-2 family, and thus, most likely, in at least partial prevention of apoptosis. It will be of great interest to examine whether iron has the potential to prevent apoptosis in the case of oxidative stress, or whether it potentiates it, via an increased generation of \*OH. With respect to iron deprivation, most data are in favor of a proapoptotic effect, and it has been established that indeed it leads to the distinctive morphological and biochemical features of apoptosis [9, 10, 14, 15].

The phase(s) of apoptosis in which iron deprivation intervenes is not known, and should also be the subject of further studies.

## TAKING AWAY IRON: WHICH DISEASES MIGHT BENEFIT?

Human beings have evolved a critical equilibrium with iron. Iron deficiency anemia affects ≈ 20% of the world's population, and any "therapy by taking away" should be followed up with all necessary care so as not to lead to an aggravation of these numbers. In contrast, the toxic effects of iron in terms of oxidative injury are best exemplified by the pathology of patients with primary or secondary hemochromatosis. Hemochromatosis is the most frequent inherited disease in European populations: homozygosity for the hemochromatosis genes, a recessive trait that leads to iron excess in tissues, occurs in 0.3% of individuals [16], while hemochromatosis secondary to hypertransfusion is an iatrogenic model for iron overload. In both genetically determined and iatrogenic hemochromatosis, the major complication is cardiac failure. The control of iron levels, by exogenous manipulations of iron status, in particular by iron chelators such as desferrioxamine, or more recently IRCO11 [17], or by controlled iron deficiency, can prevent oxidative stress and ROS toxicity.

But the beneficial effects of iron deprivation are not restricted to situations of known iron overload, and iron deprivation also represents an efficient approach for cardio-protection in non-iron-overloaded organisms. A considerable number of studies, whether in rats, rabbits, or dogs, have established the beneficial effects of iron deprivation in ischemia-reperfusion and post-ischemic damage, whether in the brain or the heart [reviewed in Ref. 18], which is in agreement with an ROS-dependent mechanism for reperfusion injury [19].

Furthermore, beneficial effects of iron deprivation have been established or suggested in diseases as diverse as anthracycline cardiotoxicity, organ transplantation, diseases with an inflammatory component such as bleomycininduced pulmonary fibrosis, bacterial infections and malaria, and, more recently, in DMD [18, 20]. All of these diseases have an oxidative component to their pathogenesis, which suggests that iron deprivation might be beneficial in many other diseases as well. One of the most impressive studies on the beneficial effects of "taking away" reports an 85% prevention of paraplegia secondary to spinal cord injury in swine [21]. Still another example is Alzheimer disease, in which iron also represents an essential source of brain oxidative alterations [22]. Finally, the contribution of iron to the toxic effects of tobacco smoke on atherosclerosis has not been considered before, although iron may mediate a great part of the toxicity of tobacco smoke, whether carcinogenesis or cardiovascular side-effects [23].

#### IRON IN INFLAMMATION

ROS are among the major mediators of inflammation, and antioxidant therapy has been proposed in many acute or chronic inflammatory diseases, although with limited success. Interestingly, although chronic inflammatory conditions are often associated with iron-deficient anemia, nutritional iron deficiency has been reported to relieve inflammation in a number of these conditions, in particular in adjuvant-induced joint inflammation [24]. Iron-deficient diet has been shown to decrease inflammation significantly in bleomycin-induced pulmonary fibrosis in hamsters [25], a condition mediated by ROS and in particular \*OH, and by TNFα. Like anthracyclines, bleomycin has high affinity for DNA and enters iron-dependent oxidation-reduction cycles, generating high levels of OH, while at the same time activating NF-κB, as reflected by TNFα release by alveolar macrophages, thus preventing apoptosis of intrapulmonary inflammatory cells.

The outcome of inflammation strongly depends upon apoptosis of the inflammatory cells [8, 11]. Thus, since iron deprivation increases apoptosis, its beneficial effects on inflammation could depend upon dual target mechanisms: prevention of oxidative damage on the one hand, and promotion of apoptosis on the other. Iron deprivation may also exert more specific beneficial effects on the immune response leading to inflammation, such as blocking interleukin-2 receptor expression on T lymphocytes, thus controlling T cell proliferation [26].

#### IRON AND INFECTIOUS DISEASES

Iron removal may have distinct therapeutic effects in bacterial infections. Indeed, iron is an indispensable nutrient for many bacteria, to the point that bacteria have evolved mechanisms to acquire and secure iron from host cells during host-pathogen interactions [27]. Mycobacteria, whether tuberculosis or paratuberculosis, are paradigmatic for such requirements. The multiplication of the bacteria appears greatly enhanced within iron-replete macrophages, whether murine [28] or human (Boshoff T, Bachelet M, Polla BS and Bornman L, unpublished results). Iron levels of the host thus appear as an essential determinant in the outcome of infectious diseases. In humans, this has been particularly well studied for tuberculosis. Indeed, a relationship between death from tuberculosis and iron overload of the liver and mononuclear-macrophage system (the latter having the strongest association) has been established [29], based on observations from the early twentieth century on iron overload in South African blacks [30]. This condition appears to result from an interaction between the amount of dietary iron and a gene distinct from the HLA-linked iron-loading locus responsible for hereditary hemochromatosis, and has been proposed as an essential component of the increased incidence of tuberculosis in South Africa.

Further supporting a role for iron in bacterial virulence, nutritional iron deficiency attenuates Salmonella typhi-

murium infection in mice as compared with both ironsubstituted and normal diet control animals [31]. In malaria, iron deprivation might exert dual beneficial effects, by acting both on the plasmodium itself and on the syndromes caused by the infection, in which ROS-dependent mechanisms again play a key role [32]. In candidiasis, iron deprivation has been shown to restore the antifungal effector functions of phagocytic cells [33].

In the light of these data, although scarce as yet, the possibility that moderate iron deprivation may represent an additional approach for the limitation of pathogenicity and spreading of infectious diseases, including AIDS, should be carefully considered.

#### IRON DEPRIVATION IN DMD

DMD is a severe genetic disorder due to mutations in the dystrophin gene at Xp21, which prevent the expression of this cytoskeletal, membrane-associated protein in affected individuals, leading to progressive muscle wasting [34]. While the absence of dystrophin is clearly responsible for the DMD phenotype, the exact role of its deficiency in the pathobiochemistry of DMD has remained elusive. One of the approaches taken in several studies to the pathogenesis of DMD has been to consider free radical reactions. Classical antioxidant therapeutic trials, however, have been inconclusive. We thus hypothesized that iron removal may provide a useful starting point for controlling free radical reactions in DMD. This hypothesis was tested in the murine model for DMD, the mdx C57BL/10ScSn inbred mice [35], which are genetically homologous to DMD. Although the use of desferrioxamine has been proposed for DMD before [36], we tested dietary iron deprivation as the method of choice for decreasing iron levels, because chelating agents such as desferrioxamine might exert specific effects distinct from iron deprivation. The results were reported by Bornman et al. [20]: there is a significant decrease in necrotic fibers in iron-deprived mdx mice, which may imply that the iron-driven generation of OH does indeed play a role in necrosis of dystrophin-deficient muscle fibers and the consequent attraction of inflammatory cells. This reduction in muscle necrosis in the mdx mouse following iron deprivation provides a basis for new approaches in the treatment of DMD patients, which is particularly exciting because of the lack of therapy for DMD and the current failure of a timely application of gene therapy. Something as simple as iron deprivation should be examined without delay in other animal models as well as in clinical trials, in all myopathies with an oxidative component including those in which classical antioxidants have not proven efficient [37, 38].

# IRON DEPRIVATION AS AN ANTI-AGING FACTOR

Cellular, tissue, and organismal aging has been convincingly associated with a progressive oxidant/antioxidant

1348 B. S. Polla

imbalance, and decreasing oxidative stress together with increasing antioxidant defenses has provided the source for most *fountains of youth*.

The increased longevity of women as compared with men has been proposed to be related to behavioral components such as a lower exposure to sources of oxidants, including tobacco smoke. I would like to propose here that this evolutionary advantage rather relates to chronic iron deficiency in women during their reproductive life, secondary to menstruation, pregnancies, and deliveries. If indeed oxidative stress and progressive oxidant/antioxidant imbalance is a major component of aging, then iron deprivation might prevent much of the cellular, tissue, and organismal lesions associated with aging and contribute to successful aging. In this perspective, it would be of great interest to examine the iron status of centenarians, along with other indicators of successful aging [39].

Skin aging, a most efficient witness of general premature aging, also presents as an oxidant/antioxidant imbalance, favored by UV radiation exposure [40]. Iron is involved in UV-induced connective tissue degradation and DNA damage [41, 42], and the possibility that moderate iron deprivation might protect skin from UV-induced premature aging and cancer should thus be considered both in experimental animal models and in human cosmetic pharmacotherapy.

If our hypothesis holds true, moderate iron deprivation might shortly add to other anti-aging "miracles" such as fruits and vegetables, supplementation in vitamin E, selenium, and others, and increase individual potential for successful aging, whether skin or organismal aging.

#### **CONCLUSION**

In conclusion, therapy by taking away (iron) has a great potential for many very different diseases, all of which share ROS-mediated mechanisms. The development of new, non-toxic, easily administrable iron chelators such as IRCO11 may shortly become the most efficient and fashionable antioxidant, anti-aging, anti-infectious, and anti-inflammatory therapy. In the meantime, although taking away by controlled dietary deprivation is less attractive, it should be considered in all of the above, as well as in the currently incurable, devastating genetic or acquired myopathies such as DMD.

I am grateful to Dr. Liza Bornman, Ph.D., Johannesburg, for inspiration and for bringing to my attention the thesis of A. S. Strachan; to Dr. Marie-Jeanne Richard, Ph.D., Grenoble, for stimulating discussions; and to Dr. Maria Bachelet Ph.D., Paris, for critical review.

#### References

- Halliwell B and Gutteridge JMC, Role of free radicals and catalytic metal ions in human disease: An overview. Methods Enzymol 86: 1–85, 1990.
- 2. Rouault TA and Klausner RD, The impact of oxidative stress

- on eukaryotic iron metabolism. In: Stress-Inducible Cellular Responses (Eds. Feige U, Morimoto RI, Yahara I and Polla BS), pp. 183–197. Birkhäuser, Basel, 1996.
- 3. Rouault TA and Klausner RD, Regulation of iron metabolism in eukaryotes. Curr Top Cell Regul 35: 1–19, 1997.
- Bornman L, Baladi S, Richard MJ, Tyrrell R and Polla BS, Differential regulation and expression of stress proteins and ferritin in human monocytes. J Cell Physiol 178: 1–8, 1999.
- Oberle S, Polte T, Abate A, Podhaisky HP and Schroder H, Aspirin increases ferritin synthesis in endothelial cells: A novel antioxidant pathway. Circ Res 82: 1016–1020, 1998.
- Cope FO and Wille JJ, Carcinogenesis and apoptosis: Paradigms and paradoxes in cell cycle and differentiation. In:
   Apoptosis: The Molecular Basis of Cell Death (Eds. Tomei LD and Cope FO), pp. 61–86. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1991.
- Zychlinsky A and Sansonetti P, Apoptosis as a proinflammatory event: What can we learn from bacteria-induced cell death? Trends Microbiol 5: 201–204, 1997.
- Polla BS, Banzet N, Dall'Ava J, Arrigo AP and Vignola AM, Les mitochondries, carrefour entre vie et mort cellulaire: Rôles des HSP et conséquences sur l'inflammation. Médecine/ Sciences 14: 1–8, 1998.
- Kroemer G, Petit P, Zamzami N, Vayssière JL and Mignotte B, The biochemistry of programmed cell death. FASEB J 9: 1277–1287, 1995.
- Leist M and Nicotera P, The shape of cell death. Biochem Biophys Res Commun 236: 1–9, 1997.
- Vayssier M, Banzet N, Bellmann K, François D and Polla BS, Tobacco smoke induces both apoptosis and necrosis in mammalian cells: Differential effects of Hsp70. Am J Physiol 275: L771–L779, 1998.
- 12. Buttke TM and Sandstrom PA, Oxidative stress as a mediator of apoptosis. *Immunol Today* **15:** 7–10, 1994.
- Jacob AK, Hotchkiss RS, DeMeester SL, Hiramatsu M, Karl IE, Swanson PE, Cobb JP and Buchman TG, Endothelial cell apoptosis is accelerated by inorganic iron and heat via an oxygen radical dependent mechanism. Surgery 122: 243–253, 1997
- Haq RU, Werely JP and Chitambar CR, Induction of apoptosis by iron deprivation in human leukemic CCRF-CEM cells. Exp Hematol 23: 428–432, 1995.
- Kovar J, Stunz LL, Stewart BC, Kriegerbeckova L, Ashman RF and Kemp JD, Direct evidence that iron deprivation induces apoptosis in murine lymphoma 38C13. *Pathobiology* 65: 61–68, 1997.
- Cox TM and Kelly AL, Hemochromatosis: An inherited metal and toxicity syndrome. Curr Opin Genet Dev 8: 274– 281, 1998.
- 17. Rivkin G, Link G, Simhon E, Cyjon RL, Klein JY and Hershko C, IRCO11, a new synthetic chelator with selective interaction with catabolic red blood cell iron: Evaluation in hypertransfused rats with hepatocellular and reticuloendothelial radioiron probes and iron-loaded rat heart cells in culture. Blood 90: 4180–4187, 1997.
- 18. Hershko C, Control of disease by selective iron depletion: A novel therapeutic strategy utilizing iron chelators. *Baillieres Clin Haematol* 7: 965–1000, 1994.
- 19. McCord JM, Oxygen-derived free radicals in postischemic tissue injury. N Engl J Med 312: 159–163, 1985.
- Bornman L, Rossouw H, Gericke GS and Polla BS, Effects of iron deprivation on the pathology and stress protein expression in murine x-linked muscular dystrophy. *Biochem Pharma*col 56: 751–757, 1998.
- Qayumi AK, Janusz MT, Jamieson WR and Lyster DM, Pharmacological interventions for prevention of spinal cord injury caused by aortic crossclamping. J Thorac Cardiovasc Surg 104: 256–261, 1992.

- Smith MA, Harris PLR, Sayre LM and Perry G, Iron accumulation in Alzheimer disease is a source of redox-generated free radicals. Proc Natl Acad Sci USA 94: 9866–9868, 1997.
- Wesselius LJ, Nelson ME and Skikne BS, Increased release of ferritin and iron by iron-loaded alveolar macrophages in cigarette smokers. Am J Respir Crit Care Med 150: 690–695, 1994.
- Andrews FJ, Morris CJ, Lewis EJ and Blake DR, Effect of nutritional iron deficiency on acute and chronic inflammation. Ann Rheum Dis 46: 859–865, 1987.
- Chandler DB, Barton JC, Briggs DD, Butler TW, Kennedy JI, Grizzle WE and Fulmer JD, Effects of iron deficiency on bleomycin-induced lung fibrosis in the hamster. Am Rev Respir Dis 137: 85–89, 1988.
- Carotenuto P, Pontesilli O, Cambier JC and Hayward AR, Desferoxamine blocks IL 2 receptor expression on human T lymphocytes. J Immunol 136: 2342–2347, 1986.
- 27. Weinberg ED, Patho-ecological implications of microbial acquisition of host iron. Rev Med Microbiol 9: 171–178, 1998.
- Lepper AWD, Jarrett RG and Lewis VM, The effect of different levels of iron intake on the multiplication of Mycobacterium paratuberculosis in C57 and C3H mice. Vet Microbiol 16: 369–383, 1988.
- 29. Gordeuk VR, McLaren CE, MacPhail AP, Deichsel G and Bothwell TH, Associations of iron overload in Africa with hepatocellular carcinoma and tuberculosis: Strachan's 1929 thesis revisited. *Blood* 87: 3470–3476, 1996.
- Strachan AS, Haemosiderosis and haemochromatosis in South African natives with a comment on etiology of haemochromatosis.
   M.D. Thesis. University of Glasgow, Glasgow, UK, 1929.
- Puschmann M and Ganzoni AM, Increased resistance of iron-deficient mice to Salmonella infection. Infect Immun 17: 663–664, 1977.
- Harvey PWJ, Bell RG and Nesheim MC, Iron deficiency protects inbred mice against infection with *Plasmodium* chabaudi. Infect Immun 50: 932–934, 1985.
- 33. Mencacci Á, Cenci E, Boelaert JR, Bucci P, Mosci P, Fe d'Ostiani C, Bistoni F and Romani L, Iron overload alters

- innate and T helper cell responses to Candida albicans in mice. J Infect Dis 175: 1467–1476, 1997.
- Engel AG, Duchenne dystrophy. In: Myology (Eds. Engel AG and Banker BQ), pp. 1185–1240. McGraw-Hill, New York, 1986.
- 35. Partridge TA, Morgan JE and Coulton GR, The mdx mouse: A model of Duchenne muscular dystrophy. In: Duchenne Muscular Dystrophy Animal Models and Genetic Manipulation (Eds. Kakulas BA, MacHowell J and Roses AD), pp. 95–104. Raven Press, New York, 1992.
- Clark IA, Proposed treatment of Duchenne muscular dystrophy with desferrioxamine. Med Hypotheses 13: 153–160, 1984.
- Jackson MJ and Edwards RHT, Free radicals and trials of antioxidant therapy in muscle diseases. Adv Exp Med Biol 264: 485–491, 1990.
- 38. Austin L, de Nieses M, McGregor A, Arthur H, Gurusinghe A and Gould MK, Potential oxy-radical damage and energy status in individual muscle fibers from degenerating muscle diseases. *Neuromuscul Disord* 2: 27–33, 1992.
- Franceschi C, Montio D, Sansoni P and Cossarizza A, The immunology of exceptional individuals: The lesson of centenarians. *Immunol Today* 16: 12–16, 1995.
- Mariéthoz E, Richard MJ, Polla LL, Kreps S, Dall'Ava J and Polla BS, Oxidant/antioxidant imbalance in skin aging: Environmental and adaptative factors. *Rev Environ Health* 13: 147–168, 1998.
- 41. Brenneisen P, Wenk J, Klotz LO, Wlaschek M, Briviba K, Krieg T, Sies H and Scharffetter-Kochanek K, Central role of ferrous/ferric iron in the ultraviolet B irradiation-mediated signaling pathway leading to increased interstitial collagenase (matrix-degrading metalloprotease (MMP)-1) and stromely-sin-1 (MMP-3) mRNA levels in cultured human dermal fibroblasts. J Biol Chem 273: 5279–5287, 1998.
- Cai CX, Birk DE and Linsenmayer TF, Nuclear ferritin protects DNA from UV damage in corneal epithelial cells. Mol Biol Cell 9: 1037–1051, 1998.